

FASTQ QC Report

Report Date	10-02-16
Run ID	160930_D00796_0121_AC9MR4ANXX
Project ID	EC-EL-3883
Sample	Sample_OD3_R1
FASTX-Toolkit Version	0.0.13.2
FastQC Version	0.10.1
Dupest Version	0.1.0

This report was automatically generated by the WCMC Epigenomics Core QC pipeline and contains information for assessing the quality of FASTQ sequencing data.

The QC Pipeline executes the following analysis:

1. All FASTQ files for the sample are concatenated to a single file. For paired-end sequences, FASTQ files for each read are concatenated and processed separately, with an "R1" or "R2" appended to the sample name.
2. To identify genomic sequencing bias or low sequence diversity k-length oligonucleotide enrichment is calculated and plotted from the combined FASTQ file using FastQC. *Note:* FastQC only analyses the top 2% of the reads in the FASTQ file and the results are extrapolated over the remainder.
3. Duplication level is estimated from the combined FASTQ file as $(N - U)/N$ where N is total reads and U is the number of unique sequences.
4. Sequencing base call quality statistics are calculated from the combined FASTQ file using FASTX-Toolkit FASTQ Quality Filter.

The report contains the following figures:

1. Sequence Duplication - Estimate of duplication level as a percentage of total reads.
2. Base sequence quality - Calculated from FASTX-Toolkit FASTQ Quality Filter.

Distribution of base quality scores (Q scores) per sequencing cycle. In a reasonably good sequencing run the majority of the signal should be above Q30. Quality scores are divided into three ranges: green indicates calls of very good quality; orange indicates calls of reasonable quality and red indicates calls of poor quality.

Yellow boxes represent the inter-quartile range. Upper and lower whiskers represent the maximum and minimum excluding outliers. The red line represents the median quality and the blue line represents the mean quality.

3. Sequence base content - Percentage of bases represented at each position in the read; calculated from FASTX-Toolkit FASTQ Quality Filter.
4. K-mer content - calculated and plotted by FastQC. From FastQC Help:

The k-mer analysis checks if there are short fragments of k-length nucleotides that are over represented at certain positions in the reads. In a diversified library there should not be positional bias in its appearance of k-mers. There may be biological reasons why certain k-mers are enriched or depleted overall, but these biases should affect all positions within a sequence equally. In contrast, if certain k-mers are over represented in certain positions then this could indicate issues with library preparation, quality of the input material or sequencing of the adaptors. This analysis measures the number of each 5-mer at each position in the library and then uses a binomial test to look for significant deviations from an even coverage at all positions. Any k-mer with positionally biased enrichment are reported. The top 6 most biased k-mers are additionally plotted to show their distribution. Note that because of the computational overhead associated with calculating k-mer content this analysis is performed on 2% of the reads.

5. Overrepresented sequences - Calculated and plotted by FastQC. From FastQC Help:

A normal high-throughput library will contain a diverse set of sequences, with no individual sequence making up a tiny fraction of the whole. Finding that a single sequence is very overrepresented in the set either means that it is highly biologically significant, or indicates that the library is contaminated, or not as diverse as you expected.

This analysis lists all of the reads which make up more than 0.1% of the total. To limit memory use only sequences which appear in the first 200,000 sequences are evaluated for their occurrences in the entire library. It is possible that a sequence which is overrepresented but doesn't appear at the start of the file for some reason

could be missed by this analysis. However, this is unlikely since library preparation and sequencing randomize the genomic elements and therefore the first 200,000 reads are sufficient to represent the diversity in the entire library.

For each overrepresented sequence the program will look for matches in a database of common contaminants and will report the best hit it finds. Hits must be at least 20bp in length and have no more than 1 mismatch. Finding a hit doesn't necessarily mean that this is the source of the contamination, but may provide clues about the true source of contamination. It's also worth pointing out that many adapter sequences are similar in sequence so a match to an adaptor sequence may not represent the true source of the adaptor.

Because the duplication detection requires an exact sequence match over the whole length of the sequence. Reads over 75bp in length are truncated to 50bp for the purposes of this analysis.

FastQC: <http://www.bioinformatics.babraham.ac.uk/projects/fastqc>

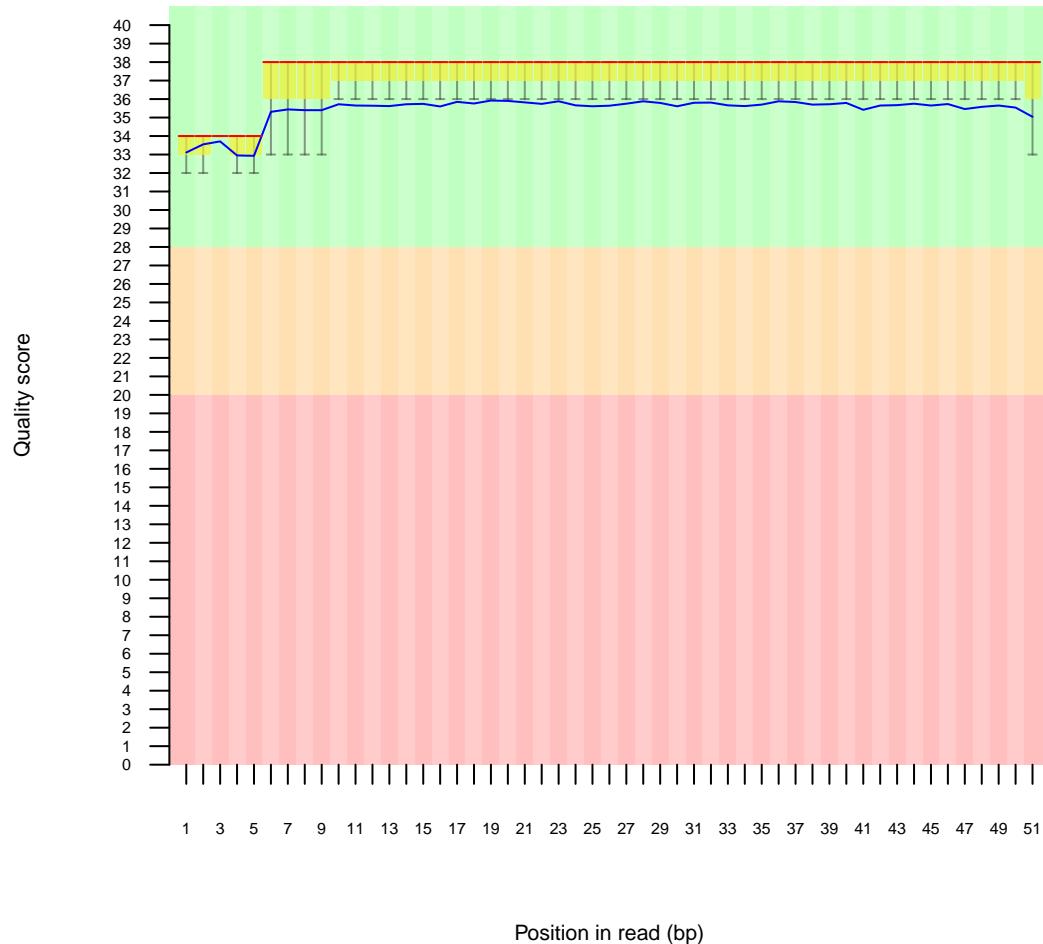
FASTX-Toolkit: http://hannonlab.cshl.edu/fastx_toolkit

1 Sequence Duplication

- Estimated Duplication rate 76.2011%

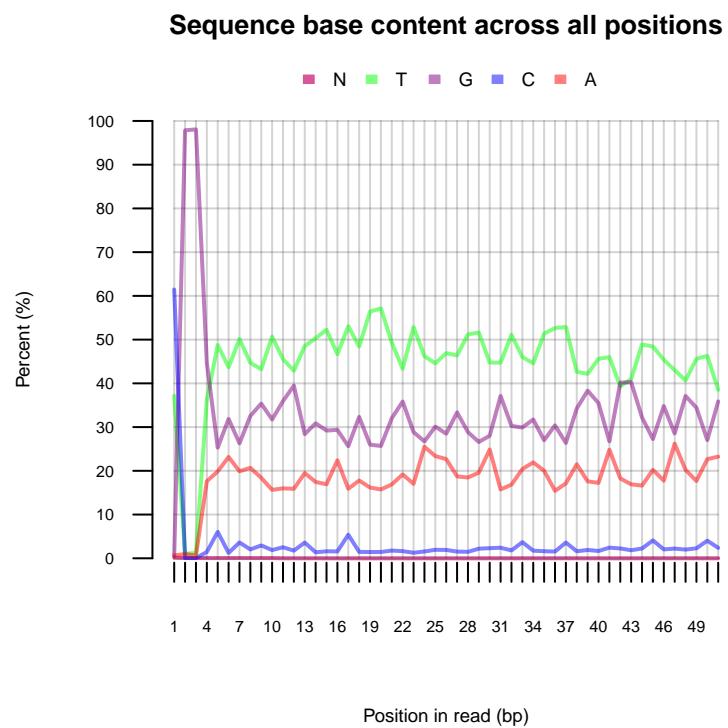
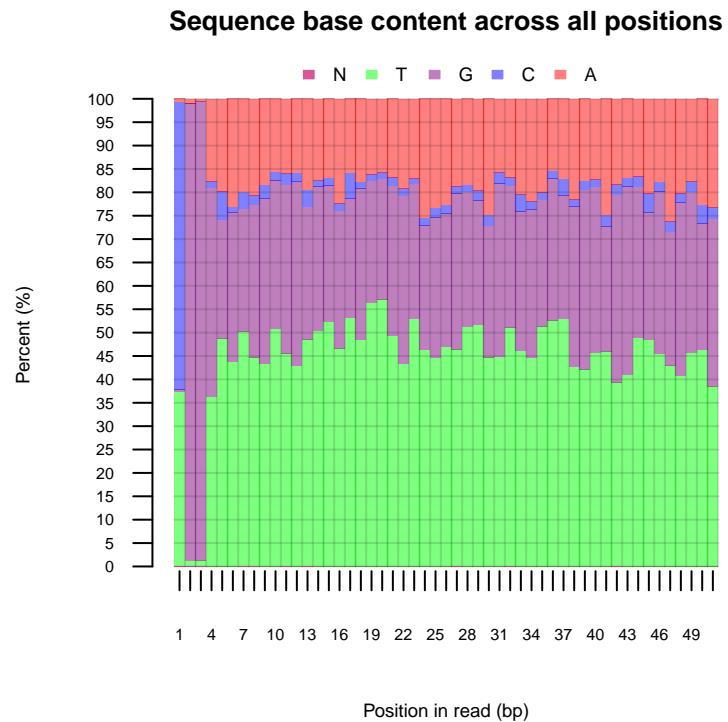
2 Per base sequence quality

Quality scores across all bases

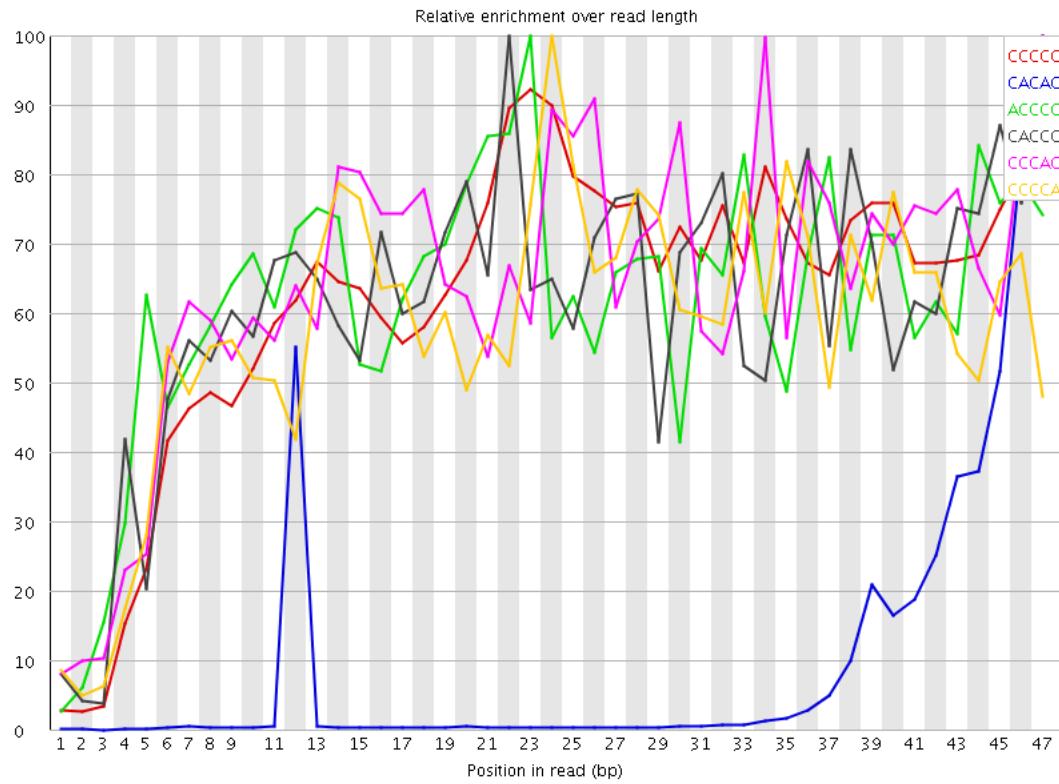


Background colors	Green - calls of very good quality Orange - calls of reasonable quality Red - calls of poor quality
Yellow boxes	Inter-quartile range
Upper and lower whiskers	Maximum and minimum quality excluding outliers
Red line	Median quality
Blue line	Mean quality

3 Sequence base content



4 Sequence K-mer content



Note: FastQC analyses 2% of the sequence data and results are extrapolated to the rest of the sequence.

Sequence	Count	Obs/Exp Overall	Obs/Exp Max	Max Obs/Exp Position
CCCCC	114430	830.5456	1321.7344	47
CACAC	513875	125.43485	1219.104	47
ACCCC	42190	56.156715	90.07055	23
CACCC	40990	54.559464	88.50682	22
CCCCAC	40565	53.99377	83.50689	47
CCCCA	40445	53.834045	92.26018	24
CCACC	39980	53.21511	77.2513	35
CGGGC	4070650	28.873442	1051.5446	1
GCCCC	24805	17.866243	34.3565	47
AGCAC	722260	17.495436	126.10145	45
CCCGC	23240	16.739021	29.956036	34
CGGCC	23010	16.573362	30.463478	32
CCGCC	22885	16.483328	29.447567	27
CCCGG	22490	16.198822	30.462605	25
GCACA	621040	15.043563	125.94261	46
CGCGG	1781095	12.633447	317.5193	5
GCGCG	1681260	11.925309	315.72055	4
CGGAA	4269965	10.264216	212.09207	1
CGGCG	1372665	9.736421	272.9318	1
ACACC	396960	9.615634	101.28569	47
GGCAC	1298460	9.210079	316.35666	3
CGCGC	127845	9.137937	56.245094	13
CTCCC	16795	9.030739	14.276814	29
CCTCC	16655	8.955461	13.897709	28
TCCCC	16400	8.818346	16.94055	3
CCCTC	14945	8.035986	13.013034	22
CCCTT	14620	7.8612323	12.129037	38
CGGGA	6007210	7.8140497	240.73703	1
ACTCC	70045	6.9069924	256.645312	23
CTCCA	69235	6.8271203	256.64578	24
CGGGT	11890205	6.2480397	244.70288	1
AGACG	2577315	6.1953955	73.35956	27
TCGCG	1118780	5.9241867	26.602547	30
CGGAG	4368605	5.6825867	170.68391	1
AGATC	3065705	5.501501	26.57111	43
CGGTT	13145150	5.156669	182.91411	1
ACGCG	391915	5.1371803	14.583303	14
AACCC	20660	5.0430236	8.201811	32
CGCGT	935560	4.9539967	24.654167	31
CGGGG	6879860	4.842669	116.88244	1
CCACA	19575	4.77818	7.513342	15
ACACC	19545	4.770858	7.9147534	21
CGGAC	357610	4.6875143	157.3611	1

CGATC	80695	0.78963953	5.0148044	44
GGTAA	4413635	0.78599113	6.5336685	2
GTCG	1467730	0.7712597	6.65729	4
TGGGT	19736735	0.7683329	8.953083	1
AGTGG	7881830	0.75953907	5.293362	8
GTTGG	19281560	0.75061333	5.20816	39
CGAAC	310015	0.7452195	6.430682	2
TGGTG	18656105	0.72626495	5.688867	7
GAGTC	710330	0.6897835	13.45698	21
TCCAG	69505	0.68014	26.535437	25
ATGCC	68535	0.6706479	28.649637	47
CAGTC	66840	0.6540617	26.705927	27
GCATC	66645	0.6521535	27.554714	38
TGGTA	9016850	0.64867496	5.175776	1
GTCAC	66220	0.64799464	27.271757	29
CCAGT	65980	0.6456461	26.452816	26
ATCTC	71835	0.5247683	21.284435	40
TGGAT	7100760	0.51083094	5.098984	1
CATCT	68615	0.50124556	20.53947	39
GATTC	687245	0.4982112	5.346739	29
CACTT	67410	0.49244285	19.871918	31
TCACT	66095	0.48283648	20.028067	30
GGTGC	787000	0.4135511	6.718141	3
TGAAC	205100	0.36805817	5.309803	20
TCTCG	76870	0.30387166	11.563231	41
CTCGT	76250	0.30142078	11.579086	42
CTGAA	137920	0.24750163	5.065655	19
AGGCA	83535	0.2008029	6.8378277	36
GAACT	95055	0.17057909	5.065238	21
AGTCA	75310	0.13514608	5.053123	28

5 Overrepresented sequences

Note: FastQC tracks sequences that appear in the first 200,000 reads to the end of the file.

Sequence	Count	%	Possible Source
CGGGTTACGTTATTTTTGTAGTTTCGAGTAGTTGGGATTATAG	198718	0.2776040174086516	No Hit
CGGGCGCGTGGTTACGTTGTAATTAGTATTTGGAGGTCGAGGCG	186185	0.26009573355825744	No Hit
CGGGTTACGTTATTTTTGTAGTTTAAGTAGTTGGGATTATAG	120927	0.1689319589225738	No Hit
CGGTTAATTGTAGAGACGGGTTATCGTGTAGTTA	87553	0.12230932545707823	No Hit